Sulforaphane improves cognitive function administered following traumatic brain injury

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Abstract

Recent studies have shown that sulforaphane, a naturally occurring compound that is found in cruciferous vegetables, offers cellular protection in several models of brain injury. When administered following traumatic brain injury (TBI), sulforaphane has been demonstrated to attenuate blood-brain barrier permeability and reduce cerebral edema. These beneficial effects of sulforaphane have been shown to involve induction of a group of cytoprotective, Nrf2-driven genes, whose protein products include free radical scavenging and detoxifying enzymes. However, the influence of sulforaphane on post-injury cognitive deficits has not been examined. In this study, we examined if sulforaphane, when administered following cortical impact injury, can improve the performance of rats tested in hippocampal- and prefrontal cortex-dependent tasks. Our results indicate that sulforaphane treatment improves performance in the Morris water maze task (as indicated by decreased latencies during learning and platform localization during a probe trial) and reduces working memory dysfunction (tested using the delayed match-to-place task). These behavioral improvements were only observed when the treatment was initiated 1 h, but not 6 h, post-injury. These studies support the use of sulforaphane in the treatment of TBI, and extend the previously observed protective effects to include enhanced cognition.

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Cells possess endogenous protective mechanism(s) as a defense against environmental insults and injury. These cellular defenses often involve induction of cytoprotective genes whose protein products scavenge and/or neutralize toxins. A few regulatory pathways have been identified involving specific transcription factors that induce the expression of cytoprotective proteins and enzymes [11]. The transcription factor Nrf2 (nuclear factor E2-related factor 2) binds to the antioxidant/electrophilic response element (ARE/EpRE) and regulates the expression of multiple cytoprotective proteins including antioxidant and glutathione generating enzymes [8–15]. Nrf2 is normally sequestered in the cytoplasm through its interaction with KEAP1 where it is rapidly turned-over by the ubiquitin-proteasome system [9–16]. The naturally occurring compound sulforaphane, which is present in cruciferous vegetables such as broccoli, has been shown to reduce the proteosomal degradation of Nrf2, resulting in its nuclear accumulation and increased expression of Nrf2-driven genes [16].

Our previous studies have shown that systemic administration of sulforaphane results in increased expression of Nrf2-driven genes in rodent brains and microvasculature [17,18]. Consistent with this, post-injury administration of this compound to rodents with traumatic brain injury (TBI) attenuates blood-brain barrier permeability and reduces brain edema [17,18]. In the present study, we investigated if post-injury sulforaphane administration improves cognitive function. Our results indicate that sulforaphane administration initiated at 1 h, but not at 6 h, post-cortical impact injury improves spatial learning and memory, and reduces working memory dysfunction.

Male Sprague Dawley rats (300 g) were purchased from Harlan (Indianapolis, IN). Sulforaphane (Catalog No. S8044) was purchased from LKT Laboratories, Inc. (St. Paul, MN). Anti-4-Hydroxy-2-noneal (4-HNE) antibody was purchased from EMD Chemicals, Inc. (Gibbstown, NJ).

All experimental procedures were approved by the Institutional Animal Care and Use Committee and conducted in accordance with the recommendations provided in the Guide for the Care and Use of Laboratory Animals. A controlled cortical impact (CCI) device was used to cause brain injury as previously described [3]. Bilateral craniectomies were positioned midway between the bregma and lambda and animals received a single impact (2.7 mm deformation, 6 m/s) on the right parietal lobe. Sulforaphane was dissolved in corn oil and administered by intraperitoneal (i.p.) injection.

Spatial memory was tested using the standard hidden platform version of the Morris water maze task (MWM) (beginning on day 14 post-injury) followed by a working memory task (beginning on day 25 post-injury). Animals were randomly assigned to either drug...
or vehicle groups. For the MWM task, animals were trained for 7 consecutive days and were given 4 training trials (60 s maximum) per day, with an inter-trial interval (iti) of 4 min [2–5]. If the animal failed to locate the platform within 60 s on any given trial, it was led there by the experimenter where it was allowed to rest for 30 s. If no significant differences were detected during the first 5 days, training was discontinued. Twenty-four hours following the last day of training, animals were tested for retention by administering a 60 s probe trial. Movement within the maze was monitored using a video camera linked to tracking software (Ethovision, Noldus Information Technology, Leesbury, VA, USA). The time to cross the site at which the platform was located during training, the number of platform crossings, and quadrant preference was calculated.

Following MWM training and testing, animals were tested in a delayed match-to-place spatial working memory task [6]. For each testing pair of trials, animals were given a location trial, in which the platform and start location were randomized. Animals were then allowed to search for the platform for a period of 60 s (location trial) and once found, the animal was allowed to rest on it for 10 s. The animal was removed from the water maze and after a 5 s delay, allowed to once again search for the hidden platform (match trial). This procedure of location and match trial was repeated five times (each pair separated by a 4 min iti) with the platform in a novel location for each pair.

Sulforaphane (5 mg/kg) or vehicle was injected i.p. 1 h and 24 h after injury. One hour following the last injection, animals were deeply anesthetized with sodium pentobarbital (100 mg/kg) and transcardially perfused with PBS followed by 4% paraformaldehyde. Brains were removed, and coronal sections prepared on a cryostat. Sections were incubated with primary antibodies (anti-4-HNE or anti-NeuN) in 2.5% goat serum in Tris buffered saline containing 0.1% triton X-100 for 48 h at 4 °C. Immunoreactivity was detected by species-specific secondary antibodies conjugated to Alexafluors. Photomicrographs (2 sections/animal) were taken of the cortex and the dorsal hippocampus for quantification of 4-HNE immunoreactivity. Camera settings remained constant for all animals. NeuN immunoreactivity was used to define hippocampal subfields. Immunoreactivity was assessed using Image J (available from NIH) and calculated as optical density/mm².

For evaluation of behavioral data, a two-way repeated measures analysis of variance was utilized to determine group differences. A Holm-Sidak method for multiple comparisons was used to determine data points with significant differences. Optical density measurements for immunohistochemistry were evaluated using a Student’s t-test for unpaired variables. Data were considered significant at p < 0.05 and presented as Mean ± Standard Error of the Mean (S.E.M.).

We have previously demonstrated that sulforaphane given 6 h post-injury significantly reduces post-TBI blood-brain barrier permeability and cerebral edema [17,18]. To test if this administration routine is associated with a cognitive improvement, rats were injured and injected with either sulforaphane (5 mg/kg) or vehicle 6 h post-injury (n = 10/group) (Fig. 1A). Beginning on day 14 post-injury, animals were tested in the standard hidden platform version of the Morris water maze. Fig. 1A shows that there was no significant difference in water maze performance between the sulforaphane- and vehicle-treated animals when the compound was administered 6 h post-injury (group main comparison: F(1,9) = 0.499, p = 0.499).

To prolong the effectiveness of the drug, injured animals (n = 10/group) were given a dose of sulforaphane (5 mg/kg) or vehicle at 6 h post-injury, followed by a second injection at 24 h post-injury. Fig. 1B shows that the addition of a second sulforaphane injection also had no significant effect on water maze performance (group main comparison: F(1–9) = 0.603, p = 0.457). In contrast, when the drug was administered earlier, injured animals receiving sulforaphane (n = 9) at 1 h and 24 h post-injury
performed significantly better than their vehicle-treated counterparts (n = 10) (group main effect: \(F_{(1-8)} = 6.750, p = 0.032\). As these rats had significantly improved performance after 5 of training, they were given an additional 2 days of training to ensure that the differences in learning would be maintained (Fig. 1C; group main effect: \(F_{(1-8)} = 10.270, p = 0.013\). These differences in performance were not due to changes in swimming speed (swimming speed at end of training: vehicle, 28.10 ± 1.04 cm/s; sulforaphane, 30.42 ± 1.90 cm/s, n.s.)

To determine if the improved performance in the 1 h and 24 h sulforaphane-treated animals was associated with measures of platform localization, a probe test was given 24 h following the completion of training. The representative traces shown in Fig. 2A demonstrate the initial swimming paths taken by a vehicle- and sulforaphane–treated animal during the probe trial (Fig. 2A). When analyzed for quadrant dwell times, a significant preference for the target quadrant (I) (quadrant that contained the hidden platform) was found for the sulforaphane-treated animals (Fig. 2B). In contrast, the vehicle-infused animals only had a significant difference in dwell time between the target quadrant (I) and the start quadrant (III). The probe test was further analyzed for localization differences between the groups by determining the latencies to, and number of times the animals entered, concentric rings of decreasing diameter centered on the platform location. The summary graph shown in Fig. 2C shows that while the sulforaphane-treated animals proceed from the outermost ring (4×) to the platform with minimal delay, the vehicle-injected animals had significantly longer latencies to reach the inner rings (+, interaction between group and latency: \(F_{(3,48)} = 2.949, p = 0.042\). Consistent with this, the sulforaphane-injected animals were found to cross the site of the platform location significantly more often than the vehicle-injected animals (vehicle, 1.00 ± 0.33 crossings; sulforaphane, 2.33 ± 0.44 crossings, \(p = 0.028\); Fig. 2D). These measures of platform localization were not detected in animals injected with sulforaphane at either 6 h, or 6 h and 24 h, post-injury.

Beginning on day 25 post-injury, the animals treated at 1 h and 24 h post-injury were further tested using the delayed match-to-place working memory task (Fig. 3A). Fig. 3B shows representative traces from vehicle- and sulforaphane-treated rats during the location and match trials. Vehicle-injected animals did not show any significant decline in latency between the location and match trials (location trial: 36.08 ± 5.25 s; match trial: 39.24 ± 9.63 s; \(p = 0.110\)), indicating impaired working memory. In contrast, animals receiving sulforaphane reached the platform during the match trial significantly faster than during the location trial (location trial: 41.18 ± 2.48 s; match trial: 25.94 ± 2.48 s; \(p = 0.001\)). Comparison of the two groups revealed a significant interaction between group and trial (\(F_{(1-8)} = 15.669, p = 0.004\).

Since a number of Nrf2-driven genes have antioxidant functions, we examined if the cognitive improvement observed following sulforaphane administration was associated with reduced oxidative damage in the brains of TBI animals. Animals were injured (n = 4/group) and at 1 h and 24 h post-injury, treated with either 5 mg/kg sulforaphane or vehicle. One hour after the last injection, brains were removed for immunological evaluation of 4-hydroxynonenal (4-HNE), a marker of lipid peroxidation [4]. Sections were double-labeled with anti-NeuN to identify neuronal layers. Fig. 4 shows representative images of the cerebral cortex immediately adjacent to the injury core (Fig. 4A) and dorsal hippocampus (Fig. 4B) from a TBI animal treated with vehicle and a TBI animal treated with 5 mg/kg sulforaphane. As the immunoreactivity for 4-HNE appeared primarily in the CA3 (an area highly vulnerable to TBI) and dentate gyrus subfields of the hippocampus in vehicle-treated animals, these areas were chosen for quantification. Fig. 4C shows that 4-HNE immunoreactivity (calculated as optical density/mm²) was significantly reduced.
Fig. 3. Post-injury administration of sulforaphane improves working memory. (A) Schematic diagram showing the protocol of delayed match-to-place testing consisting of location (Loc) and match trials, separated by a 5 s delay. Five pairs of location and match trials were performed, with the platform position in a novel site for each pair. Each pair of trials was separated by a 4 min inter-trial interval. (B) Representative traces of a vehicle and a sulforaphane (Sul)-treated animal during the location and match trials. (C) Summary data showing average latencies to the platform during the location and match trials. *p < 0.05 between location and match trials. Data are represented as mean ± SEM.

in the cortex of sulforaphane-treated animals (Student’s t-test: p = 0.0006). Likewise, 4-HNE immunoreactivity was found to be significantly reduced in the CA3 subfield of the hippocampus (Student’s t-test: p = 0.035). No significant difference was found in the dentate gyrus (Student’s t-test: p = 0.220), presumably due to the high degree of variability detected in the vehicle-treated animals (Fig. 4D).

To defend against exogenous toxins and harmful products of cellular respiration, cells contain a large number of antioxidant and detoxifying enzymes whose expressions are rapidly increased in response to threats. Many of these genes contain binding sites for the transcription factor Nrf2 and can therefore be regulated by the Nrf2-activating compound sulforaphane [8–15]. Our laboratory has previously shown that post-injury administration of sulforaphane

Fig. 4. Post-injury administration of sulforaphane decreases oxidative damage in the brain. Representative images showing the relative immunoreactivity of 4-HNE in an injured animal receiving vehicle, and an injured animal receiving 5 mg/kg sulforaphane 1 h and 24 h post-injury. Images were taken from the (A) Cerebral cortex immediately adjacent to the injury location, and (B) The ipsilateral hippocampus. Hippocampal subfields were identified by NeuN double-label immunohistochemistry and indicated by gray lines. Summary data for quantification of 4-HNE immunoreactivity from (C) The ipsilateral cortex adjacent to the site of injury, (D) the CA3 subfield, and (E) the dentate gyrus. *p < 0.05. Veh: vehicle; Sul: sulforaphane; DG: dentate gyrus.
improvement of cognitive function, simply improving vascular per-
behavioral improvements were only observed when sulforaphane
posed of neurons, astrocytes, oligodendrocytes and brain endothe-
the components of the neurovascular unit (a functional unit com-
required to determine the contribution of individual structures to
working memory is heavily dependent on the transient storage
the delayed match-to-place task are dependent on the hippocam-
and by improved performance in a working memory task. Although
improvement was evidenced by enhanced learning and memory,
treatment preserves neurologic function in injured animals. This
injury. Sulforaphane, a naturally occurring compound generated
from cruciferous vegetables such as broccoli, is a potent inducer of
antioxidant and detoxifying enzymes [15]. As such, this com-
pound has been suggested to provide broad protection against a
variety of cellular threats. For example, application of sulforaphane to
neuron-astrocyte co-cultures protects neurons against nonexciti-
toxic glutamate and hydrogen peroxide toxicity [7]. Consistent with this ability of sulforaphane to reduce oxidative damage, we observed that sulforaphane administered 1 h and 24 h post-injury significantly reduced 4-HNE immunoreactivity in the CA3 subfield of the hippocampus and the cortex surrounding the injury site. The CA3 subfield has been demonstrated to be vulnerable to lateral cortical impact injury, and damage to this area is thought to contribute to hippocampal-dependent learning and memory dysfunction [1].

We have previously demonstrated that a single, post-injury injection of sulforaphane is sufficient to stimulate the expression of Nrf2-dependent genes and reduce TBI-associated blood-brain barrier permeability and cerebral edema [17,18]. While encouraging, the consequences of sulforaphane on post-injury cognition were not tested. In this study, we present data to demonstrate that in addition to vascular protection, post-injury sulforaphane treatment preserves neurologic function in injured animals. This improvement was evidenced by enhanced learning and memory, and by improved performance in a working memory task. Although the systemic nature of the drug treatment paradigm employed in this study prevents ascribing these neurologic improvements to a specific brain structure, both the Morris water maze task and the delayed match-to-place task are dependent on the hippocampus, suggesting a preservation of this structure’s function. However, working memory is heavily dependent on the transient storage and manipulation of information by the prefrontal cortex. Thus, the improved performance observed in the delayed match-to-place task may also reflect a reduction of prefrontal cortex dysfunction. Future studies using targeted infusions of sulforaphane may be required to determine the contribution of individual structures to the effects we observed.

Based on our previous observations that sulforaphane could offer protection against blood-brain barrier compromise when delayed by as much as 6 h post-injury [17,18], and the premise that the components of the neurovascular unit (a functional unit composed of neurons, astrocytes, oligodendrocytes and brain endothelial cells) work in concert to maintain brain function, we anticipated that cognitive improvement would be seen when sulforaphane was administered 6 h post-injury. However, hippocampal-dependent behavioral improvements were only observed when sulforaphane treatment was initiated within 1 h of injury. The reason for the dissociation between vascular protection and cognitive improvement is not clear at present. As a number of factors are required for improvement of cognitive function, simply improving vascular per-
meability may not be sufficient. Furthermore, although oxidative damage may occur in the neurons and endothelial cells with similar time courses [4], there is evidence to suggest that neurons may be particularly sensitive to injury. For example, using the expression of hsp70 as a marker of injury, it has been shown that brief periods of ischemia results in hsp70 induction first in neurons, followed by glia, and finally in endothelial cells [14]. Therefore, sufficient oxidative damage may have occurred within neurons by 6 h post-injury to make this delay incapable of improving cognitive function. Regardless of the underlying reasons for the observed differences in time windows, our results on the behavioral and pathophysiological improvements offered by post-injury sulforaphane administration suggest that this compound is a promising therapeutic option for the treatment of brain injury patients.

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